

Principles of treatment of malignant gliomas in adults: An overview

S Ausim Azizi¹ and Curtis Miyamoto²

Departments of ¹Neurology and ²Radiation Oncology and ^{1,2}The Brain Tumor Center, Allegheny University of the Health Sciences, Broad and Vine, MS 423, Philadelphia, PA 19102, USA

The cornerstone of conventional treatments of malignant gliomas in adults has been surgical debulking, radiation therapy and chemotherapy. Almost always a combination of these treatments is used. With these conventional treatments the outcome, as measured by survival and quality of life, has remained universally dismal. Novel treatments, which are at different stages of laboratory and clinical trials, may offer a ray of hope for treatment of malignant gliomas. Development of these methods are directly related to the discoveries, over the past two decades, of cellular and molecular mechanisms involved in the genesis of brain tumors. Understanding of the mechanisms of tumor genesis may open new avenues of effective treatments for this devastating cancer.

Keywords: glioma; glioblastoma; radiotherapy; chemotherapy; gene therapy; TGF-beta

Introduction

Because of extremely short post-diagnostic survival time and universal fatality, malignant brain tumors are considered among the most devastating cancers that afflict human beings. Among the 10 000 to 15 000 newly diagnosed malignant brain tumors, less than 10% survive 2 years and less than 5% live to 5 years (Levine *et al*, 1989; Shoenberg, 1983). Tumors of the central nervous system are a common cause of cancer death in young adults. Although, aggressive treatment modalities have extended the median survival from 4 months to 1 year, the survival is often associated with significant impairment in the quality of life. Furthermore, it appears that the incidence of primary brain tumors is increasing, at least in the elderly (Boring *et al*, 1994). The pathogenesis of high grade primary brain tumors is characterized by local recurrence, though multifocal gliomas do occur. Current treatment of malignant gliomas can be divided into two broad categories: (1) Conventional treatments, and (2) investigational treatments. This division is somewhat artificial, as many experimental treatments are in fact innovations in conventional treatments. Furthermore, the 'experimental treatments' often rapidly move to the mainstay of the management of brain tumors.

Conventional treatments

Surgical resection, radiation therapy, and to a lesser extent, chemotherapy have been the cornerstone of treatment for high grade gliomas. Almost always a combination of these treatments is used. Improvements in surgical techniques including intra-operative mapping of the eloquent areas of brain (Black, 1997), and most recently the use of magnetic resonance theater (MRT) (Gould and Darzi, 1997; Fried *et al*, 1996) may offer some benefits for these patients in prolonging survival by a few months. However, even with aggressive surgical treatment the rate of recurrence for malignant gliomas is greater than 90%, and interestingly, the site of recurrence is often in the resected tumor bed, i.e., the recurrence remains local (Hochberg and Pruitt, 1980). In general, post-surgical quality of life remains poor.

Although the clinical benefits from radiotherapy, as measured by length of survival, appear to be modest, it is more effective than chemotherapeutic agents tested thus far, and with relatively low short term complications (Blomgren, 1996). Recent technical modifications in radiotherapy, such as hyperfractionation, quality of radiation, stereotaxic treatment with sparing of normal tissue, and addition of radiosensitizing substances have marginally improved clinical efficacy of radiotherapy for highly malignant brain tumors. The rationale behind the current methods of radiotherapy is to confine the radiation effects to the tumor area and to take advantage of the rapid division of malignant

cells and inflict damage to malignant cell nuclei at vulnerable times in the cell cycle. The damage can be at molecular or even at karyotypic levels (Lai *et al*, 1997).

Several chemotherapeutic agents, tried for the treatment of recurrent malignant gliomas, have shown only minor clinically meaningful activity against primary malignant brain tumors. With regard to systemic chemotherapy, many issues need to be considered including penetration of the blood-brain barrier and dose limiting toxicity. Therefore, there is no consensus, as yet regarding the most effective chemotherapeutic regimen. Furthermore, the logistics of administration and undesirable peripheral side effects often prove to be prohibitive.

Radiotherapy

Radiation therapy is one of the most effective treatments for anaplastic astrocytomas and glioblastoma multiforme. These tumors are theoretically ideal for radiation therapy as they are almost always confined to one location. Different methods for treatment delivery and types of irradiation have been developed.

Mechanisms of radiation effects

Ionizing radiation is thought to produce secondary electrons, which result in damage to the cellular DNA and the cell membrane, ultimately causing cell death. Many forms of DNA damage can occur. These include double and single strand breaks, base damage and DNA-protein cross-links. The double strand breaks have the greatest correlation with human tumor cell line death (Kelland *et al*, 1988; Schwartz *et al*, 1988; Wlodek and Hittelman, 1987). The exact mechanism by which ionizing radiation destroys tumor cells more efficiently than normal cells is not clear. It is widely believed that the reason for the tumor cells' sensitivity to radiation and chemotherapy is that they replicate more quickly than the normal tissues. This is partially true, though not a full explanation, because in many normal tissues, e.g. endothelial, epithelial and hematopoietic tissues, self renewing cells replicate as quickly as tumor cells. It is likely that the anticancer agents, among other mechanisms, induce p53 dependent apoptosis, thereby leading to cell death (Gupta *et al*, 1996).

Standard fractionation

The most widely used form of treatment is external beam photon irradiation. Photons can be either X-rays (artificially produced) or gamma rays (produced from radioactive decay). Commonly radiation is delivered in multiple fractions in order to decrease toxicity to the normal tissue, while increasing tumor response.

The delivery of 180 to 200 centigray (cGy) fractions of irradiation once a day, until a certain total dose is achieved, is termed 'standard fractionation.' The time interval between fractions allows the normal (and tumor) cells to undergo repair of sublethal radiation damage. This is the first of what has been termed the four 'R's of radiation biology. These are repair of sublethal damage, reassortment, reoxygenation, and repopulation. The mechanism of repair in sublethal radiation damage is not well understood. The damage repair is possibly related to the integrity of cell cycle check-points in normal tissue, where normal check-points, e.g. p21, arrests the cell cycle in radiation damaged tissue allowing time for repair. Given the prevalence of check-point defects in tumors (Waldman *et al*, 1997), radiation damaged malignant cells may proceed through cell cycle and likely into an apoptotic pathway.

Standard radiation therapy increases survival by 3–6 months (roughly doubling the survival from surgery alone). The Brain Tumor Study Group has shown an increase of median survival from 14 weeks for surgery alone, to 36 weeks with post-operative radiation treatment. The one year survivals were 3% and 24% respectively. Traditionally, total doses of 5000–7000 cGy have been given. There has been no proven survival benefit from increased total radiation doses, i.e., over 6000 cGy.

Hyperfractionation

This is the delivery of two doses of radiation daily, separated by 4–6 h. It has the theoretical advantage of delivering a higher total dose of radiation to the tumor, while minimizing damage to the normal tissue. The use of hyperfractionation may increase local control without increasing toxicity due to late effects, which are mediated by the radiation damage to slowly dividing normal tissue (Nelson *et al*, 1993; Beck-Bornholdt *et al*, 1997). The overall length of treatment is the same as for standard fractionation. Again, the rationale behind this method of treatment is to radiate the maximum number of tumor cells at vulnerable times in the cell cycle. In addition, the time between the radiation treatments allows for the repair of sublethal damage in the normal tissue. In clinical studies involving children and adults, utilization of this technique has resulted in improved local control and survival (Wara *et al*, 1986; Nelson *et al*, 1993). No dose-response correlation was noted between hyperfractionated RT doses of 48.0 and 54.4 Gy. The median survival for anaplastic astrocytoma patients at 72.0 Gy was 49.9 months. Slightly higher toxicity was noted with doses of 80 Gy and above (Nelson *et al*, 1993).

Accelerated fractionation

Another method of radiation delivery is accelerated fractionation, which is the use of standard sized fractions given twice daily. This reduces the overall number of treatment days by half (Thames

et al, 1983; Peters *et al*, 1982; Fowler, 1984). The potential late term effects are unchanged since the total number of fractions are unchanged. Theoretically, there is an increased probability of tumor control and possibly survival because of a reduction in repopulation in rapidly proliferating tumors however, this has yet to be tested in a prospective randomized trial. The acute effects of radiation are increased which may necessitate a break in treatment. In one clinical study it was noted that the rate of disease progression during the course of treatment was much lower than for standard therapy (Werner-Wasik *et al*, 1996), though in some cases contrast enhanced tumor-negative lesions mimicking a brain tumor, consisting of necrosis and reactive gliosis, were observed on Magnetic Resonance Imaging (Van Tassel *et al*, 1995).

Brachytherapy

This method, also known as interstitial implantation, involves the direct placement of one or multiple radioactive source(s) in the area of the tumor by a minimally invasive neurosurgical procedure. This allows the delivery of high doses of radiation to the target volume while minimizing radiation exposure to surrounding structures. The radiation sources can be implanted directly into the tumor cavity either for a short time or indefinitely, depending on the desired effect. A key factor in the success of brachytherapy is the ability to deliver adequate doses of radiation to the tumor tissue with a good margin of treatment. This in turn is limited by tumor size (the larger the tumor the more difficult they are to include completely and uniformly in the treatment volume), and location, i.e., accessibility to treatment (Schupak *et al*, 1995). Clinical trials with stereotactic placement of Iodine-125 and Iridium-192 combined with chemotherapy have shown improved survival for glioblastoma (Sneed *et al*, 1997). However, other studies have shown a significant rate of failure requiring re-operation (Hopkins *et al*, 1995).

Stereotactic radiosurgery

Stereotactic irradiation is the delivery of a single fraction of high dose irradiation to a limited volume of tissue in one setting, using multiple arcs or beams from different directions. There are two conventionally used methods: One is given using the 'Gamma Knife', and the other is using a linear accelerator. The gamma knife consists of multiple collimated helmets with 210 cobalt-60 sources, while the linear accelerator utilizes multiple sweeps or arcs. Both techniques are designed to deliver radiation to a targeted focus from multiple directions, thus sparing the surrounding normal tissue (Hall *et al*, 1995). Recently, a hypofractionated stereotactic radiotherapy method was tried on patients with high grade gliomas with satisfactory palliative results (Shepherd *et al*, 1997). Three dimensional reconstruction of target areas is employed in both techniques. Basic differences in techniques are presented in Table 1.

Chemotherapy

Unfortunately, despite valiant efforts, systemic chemotherapy—as measured by survival time—has been the least effective of conventional treatments for malignant gliomas. To achieve a cure for malignant glioma via chemotherapy, the drug or a combination of drugs must be effective against tumor cells, and must be given frequently and in sufficient quantities to reach the dividing malignant cells at a particular phase in the cell cycle, while sparing the normal brain tissue. The mechanism of action of commonly used anticancer treatments at a biochemical level are reasonably well understood. The chemotherapeutic agents used for treatment of malignant gliomas have been alkylating agents, BCNU (Carmustin), CCNU (Lomustin) and Procarbazine; DNA cross linking agents, Carboplatin and Cisplatin; mitosis inhibitors, Vincristine sulfate; and more recently, topoisomerase inhibitors, which are derivatives of Camptothecin, a plant alkaloid (Matsumoto *et al*, 1995; Weingart *et al*, 1995;

Table 1 General comparison of linac based versus gamma knife stereotactic irradiation.

	<i>Linear accelerator</i>	<i>Gamma knife</i>
Source	Linear accelerator	60-Cobolt
Field arrangement	Multiple arcs and static fields	Multiple static fields
Field blocking	Multileaf collimators, cerrobend, independent jaws	Not conventionally employed
Time to treat (average)	Longer	Short
Field shaping isocenter	Blocking, multiple isocenters, addition of static fields, differential weighting of fields	Multiple isocenters
Fractionation	Commonly done	Not commonly done
Availability	Commonly available	At select centers
Collimator sizes	5–50 mm	4–18 mm
Imaging for planning	CT is mandatory. Image fusion with angiography and MRI.	MRI has been the standard. Now CT and angiography compatible

Lamond *et al*, 1996; Nakatsu *et al*, 1997; Balmaceda *et al*, 1997). Generally, for treatment of malignant gliomas – similar to the treatment of other cancers – a combination of the above drugs are administered. Furthermore, chemotherapy is almost always used as an adjunct to surgical and radiation treatments. It is known that radiation of the central nervous system disrupts the blood-brain barrier (Rubin *et al*, 1994). Therefore, combination treatments may have added advantage. Thus, radiation plus chemotherapy protocols, in a variety of configurations have been studied or are currently under study (Elliott *et al*, 1996a; Warnick, 1994; Prados *et al*, 1996; Kyritsis *et al*, 1996; Ameri *et al*, 1997; Kiu *et al*, 1995; Brandes *et al*, 1996; Fountzilias *et al*, 1997; Boiardi *et al*, 1997). Although a number of protocols have failed, studies comparing different chemotherapeutic regimens have not shown superiority of one set over the others (Fujiwara *et al*, 1995).

Systemic chemotherapy for brain tumors has the added disadvantage of inability to cross the blood-brain barrier. Only a handful of chemotherapeutic agents are capable of doing so. Studies are under way to evaluate breaching of the blood brain barrier for further penetration of chemotherapeutic agents (Elliott *et al*, 1996b). In addition, the feasibility and kinetics of superselective intraarterial application of chemotherapeutic agents, i.e., infusion into the arteries that feed the tumor, is under study (Nakagawa *et al*, 1994; Fujiwara *et al*, 1995).

A recently approved modality of treatment is the placement of chemotherapeutic agents directly into the tumor cavity after resection. This treatment involves the application of BCNU (carmustin) impregnated biodegradable wafers in the tumor bed at the time of surgical resection (Sipos *et al*, 1997). Clinical studies have shown some improvement in increased post-operative survival (Valtonen *et al*, 1997).

New and experimental treatments

A major limiting factor in conventional therapies for malignant gliomas is their non-specific nature, which causes dose limiting toxicity to normal brain tissue. Despite a recent pessimistic report on the rate of success in treatment of cancer (Bailar and Gornik, 1997), giant strides in understanding the cellular and molecular biology of cancer have been made, opening the way for newer therapeutic approaches. The experimental treatments are at different stages of laboratory and clinical trials and are too numerous to exhaustively review in this article; thus we concentrate on a few of the methods, promising for the future treatment of brain tumors. These treatments are primarily biological, and take advantage of the specific molecular and immunological properties of malignant gliomas. Most of these treatments are used in conjunction

with conventional therapies in different centers, though their feasibility, safety, and efficacy remain largely unclear.

New methods of radiotherapy

Boron-neutron capture therapy

Neutrons differ from most other forms of external beam irradiation in that they have a high linear energy transfer, i.e., they deposit larger amounts of energy per length of tissue penetrated. High Linear Energy Transfer radiations are less affected by sublethal damage repair (the repair that occurs between fractions of radiation), hypoxia, and cell cycle effects, which makes neutrons extremely effective at killing both the normal and tumor cells. This fact has greatly reduced the therapeutic ratio of neutron radiation. One method of increasing the efficacy of neutron beam capture therapy is to concentrate the radiation dose to the tumor *versus* the normal brain. To accomplish this, boron is administered to the patients prior to irradiation. Boron-a stable isotope ^{10}B -captures slow neutrons and is converted to lithium and helium atoms, releasing energy for the tumor kill. Of course, sufficient quantities of boron must accumulate in the tumor tissue to allow for greater differentiation between the tumor and normal tissue. To accomplish this, a variety of boron-containing compounds with selectivity for neoplastic cells compared to normal cells have been devised (Barth and Soloway, 1997). Some of these compounds include the use of monoclonal antibodies against epidermal growth factor receptors, bispecific antibodies (Liu *et al*, 1995) and sodium borocaptate (Yang *et al*, 1997). These have been employed in clinical studies with moderate success (Barth *et al*, 1997; Nakagawa and Hatanaka 1997).

Proton beam irradiation

Protons are positively charged particles produced by a cyclotron. Proton beam irradiation has some potential advantages due to what is termed as the Bragg peak effect. The Bragg peak effect denotes that the proton beam has a very specific range. This allows the proton beam to be used for the irradiation of tumors adjacent to critical normal structures (i.e. the brain stem or optic chiasm). The biologic effect of protons is the same as for photons (X-rays and gamma rays) or electrons, which are low Linear Energy Transfer radiations. Several institutions are utilizing this technique for treatment of high grade astrocytomas (Shrieve and Loeffler, 1995).

Immune therapy

The advent of specific monoclonal antibodies directed against cell surface molecules has allowed for the definition of a number of glial epitopes

associated with gliomas, and has opened a new era in the treatment of cancers in general, and brain tumors in particular. Antibodies raised against the neural cell adhesion molecules (Patel *et al*, 1989), epidermal-growth factor receptors (Brady *et al*, 1990) and tenascin-C (Natali *et al*, 1991) are of interest for treatment of malignant gliomas. The rationale for this treatment is that the antibodies bind to the tumor tissue and cause disruption of neoplastic cell function by blocking receptors to trophic factors and/or other epitopes. Furthermore, these monoclonal antibodies can be coupled with toxins (Press *et al*, 1986) or radioactive sources (Emrich *et al*, 1996), causing further selective destruction of tumor cells.

Monoclonal antibodies against epidermal growth factor receptors (EGFR) for treatment of brain tumors have been studied extensively and have progressed to phase II clinical trials. These receptors are overexpressed in malignant gliomas (Reifenberger *et al*, 1989), whereas their expression is low in normal brain. EGFR may have a role in oncogenesis and tumor growth (Libermann *et al*, 1985). Theoretically, blocking of these receptors could inhibit proliferation of tumor cells (Weiner, 1995). Preliminary clinical studies have shown substantial *in vivo* tumor binding and concentration of one type of these antibodies, (EMD55900), after intravenous administration of a single 200 mg dose (Faillot *et al*, 1996). In a study in this institution (Miyamoto *et al*, 1995), 60 patients with clinical and radiologic diagnosis of glioblastoma were pre-operatively treated with an average of three intravenous or intra arterial infusions of iodine 125-labeled murine anti-EGFR monoclonal antibodies. The study revealed that repeated administration of these antibodies is safe and may have some benefit in the management of primary glioblastomas, especially for those patients who do not qualify for other forms of more aggressive management (Miyamoto *et al*, 1995). A tumor-specific variant of epidermal growth factor receptor has been identified (Tsugu *et al*, 1997; Wikstrand *et al*, 1997). These aberrant receptors may be present in up to 50% of gliomas. Monoclonal antibodies against this group of receptors can be another avenue of immune therapy (Okamoto *et al*, 1996).

Interleukin-2, because of its ability to mobilize the organism's immune defenses against malignant cells, is another immunotherapeutic agent. It has been demonstrated that this cytokine stimulates the proliferation of tumor infiltrating lymphocytes *in vitro* (Lorruso *et al*, 1994). In a preliminary study of long-term survival in 19 patients after intra-cavitary introduction of interleukin-2 and lymphokine activated T-cells, it was shown that this agent may increase the post-diagnostic survival time of patients with malignant gliomas (Hayes *et al*, 1995).

Interferon beta can inhibit proliferation of glioma cells *in vitro* without a similar effect on the growth

of normal astrocytes (Harada *et al*, 1995; Nehashi *et al*, 1995; Yokoyama *et al*, 1997). Furthermore, it was demonstrated that its antiproliferative effect occurs by arresting the cell cycle specifically at the S-phase (Garrison *et al*, 1996). The implications are that interferon must be available to tumor cells during the S-phase to be effective. Preliminary studies with beta interferon in children and adults with gliomas, have indicated that this form of immunotherapy may be safe and feasible (Packer, 1996).

Another approach to the immune therapy is vaccination against tumor cells, along with the boosting of the tumor specific immunity with granulocyte-macrophage colony-stimulating-factor. A recent study (Yu *et al*, 1997) demonstrated that subcutaneous inoculation of rats with irradiated GM-CSF producing tumor cells protected the animals against subsequent tumor implantation. GM-CSF has been used in the past to reconstitute immunity subsequent to aggressive chemotherapy and bone marrow transplantation (Rampling *et al*, 1994).

Trophic factors and treatment of malignant gliomas

With the increasing understanding of the role of growth factors and their receptors in the genesis of gliomas, other avenues of treatment for this cancer may open up. One such factor is Insulin Like Growth Factor-I (IGF-I) (Sandberg-Nordqvist *et al*, 1993). This factor is secreted by neoplastic cells and may, through a feedback loop, contribute to their growth, proliferation and maintenance. It has been reported that IGF-1 modulates the epidermal growth factor mediated glial cell growth in culture (Chernausek, 1993). As discussed above, EGF receptors are overexpressed in gliomas (Tuzi *et al*, 1991). It is conceivable that IGF-1 and EGF work in concert to maintain and nourish gliomas. Trojan *et al* (1993) indicated that treatment of established brain glioblastoma with antisense IGF-1 complementary DNA that blocks the synthesis of IGF-1 resulted in regression of these tumors. However, later it was noted that tumor regression may have been due to rejection of allogeneic C₆-glioma cells implanted in the rat brain (Beutler *et al*, 1997).

Other important factors, essential for maintenance and progression of tumors, therefore a target for therapy, are a variety of angiogenic molecules that are elaborated by the tumor cells (Weidner, 1996). A number of angiogenic factors have been described and several mechanisms for their activation have been proposed (Liotta *et al*, 1991; Hanahan and Folkman, 1996). Recently, it was reported that basic fibroblast growth factor (BFGF) might be involved in the initiation of angiogenesis (Czubayko *et al*, 1997). A number of angiogenesis-inhibiting drugs have progressed to clinical trial

stages (Fine, 1995). These include Marimastat, a metalloproteinase inhibitor (Uhm *et al*, 1997), Thalidomide and the calcium channel blocker CA1. Paradoxically, an important source of angiogenesis inhibitor molecules may be the neoplastic tissue itself (Good *et al*, 1990).

Transforming growth factors

Another class of trophic factors which are important in possible regression and/or maintenance of brain tumors are the Transforming Growth Factors. These families of factors act through a serine/threonin kinase membrane receptor group. Intracellular transduction is accomplished via a group of proteins (SMADS), which are immediately translocated to the cell nucleus. A number of studies have indicated that Transforming Growth Factors may contribute to apoptotic death in certain types of cells (Ohta *et al*, 1997; Tachibana *et al*, 1997). TGF- β was one of the first factors identified from malignant gliomas to suppress the immune system (Bodmer *et al*, 1989).

Recently, we have carried out a preliminary set of experiments designed to study the localization and expression of Transforming Growth Factors (TGF- β) in normal glial cell and malignant glioma cells. We utilized immunocytochemistry to detect TGF- β in

cultures of normal astrocytes, glioblastoma cells obtained from patients, and C6 glioma cells lines. A monoclonal antibody against TGF- β 1 (R&D system, Minneapolis, MN) was used. The antibodies were visualized utilizing indirect immunofluorescence technique. It was noted that in normal glial cells, TGF- β was concentrated in the cytoplasm, whereas in both glioblastoma cells and in the C6 glioma cell lines, this factor showed reactivity in the nucleus (Figure 1). The implication of these findings is not clear at this point.

Gene therapy

Advances in molecular biology in the last decade have better illustrated the mechanisms involved in the genesis of malignant gliomas. It is now generally understood that, to some extent, tumor genesis occurs either by overexpression of oncogenes or inactivation of tumor suppressor genes. Understanding the genetic and molecular mechanisms of oncogenesis represents the ultimate challenge, and likely is the only route to cure and control brain tumors.

One method of gene therapy may be the correction of genetic defect(s) either by introducing the

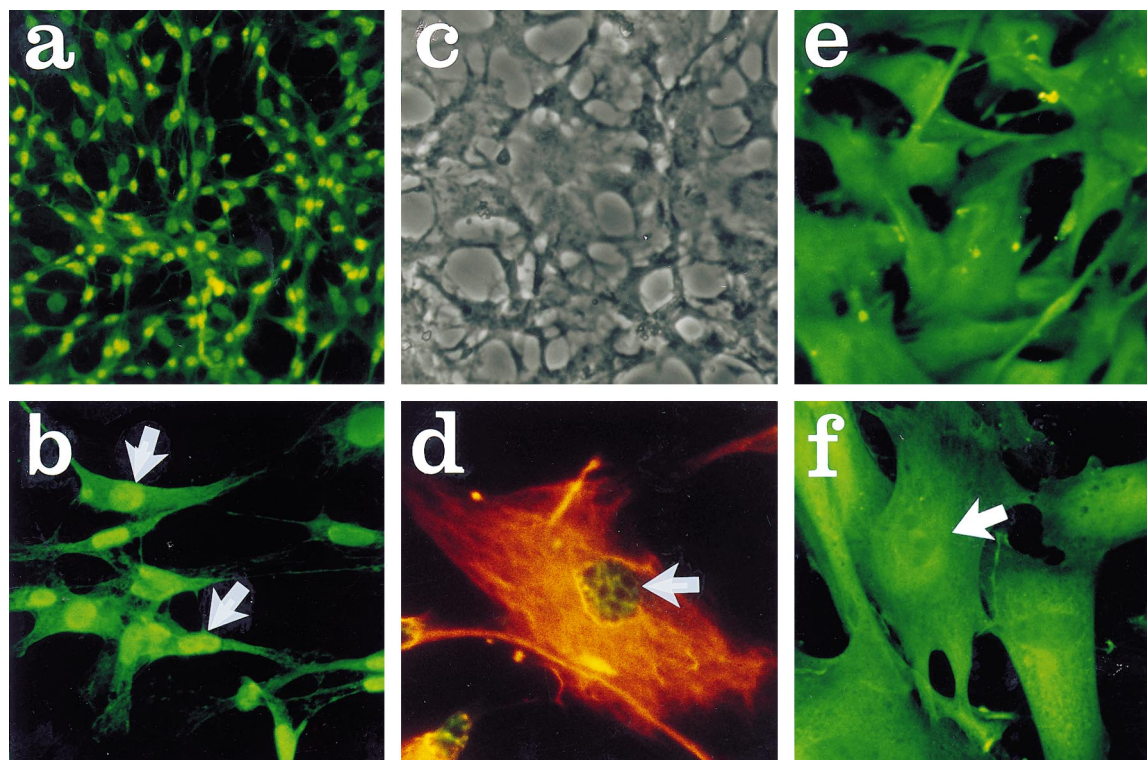


Figure 1 Photomicrographs demonstrate the localization of antibodies to TGF- β in cultures of normal and malignant glial cells. (a, b) Low and high magnification photos demonstrate concentration of TGF- β antibodies (arrowheads) in the nucleus of C6-glioma cells. (c, d) Similar findings indicate concentration of fluorescent labeled antibodies to TGF- β in the nucleus (arrow) of a primary culture of human glioblastoma, obtained from a patient. The cells were also labeled with antibodies against glial fibrillary acidic protein (GFAP), shown in orange in this double labeled section. (e, f) Indirect fluorescence immunocytochemistry indicate concentration of antibodies mainly in the cytoplasm of normal astrocytes, obtained from normal adult human brain.

missing tumor suppressor gene or by blocking the overexpression of oncogene(s). However lack of appropriate methods of gene engineering, as well as sparse understanding of the location and sequences of the gene(s) and the exact mechanisms of activation and translation, have thus far limited this approach.

A second gene therapy approach for the treatment of malignant gliomas involves the use of suicide genes such as thymidine kinase or cytosine deaminase from herpes simplex virus and *E. coli*, respectively (Ezzedin *et al*, 1991; Oldfield *et al*, 1993; Izquierdo *et al*, 1996; Kruse *et al*, 1997). This strategy can be effective either alone or by conferring chemosensitivity and radiosensitivity (Kim *et al*, 1997) to the malignant cells. Thymidine Kinase, once expressed in the replicating tumor cells, rapidly phosphorylates the antiviral agents, acyclovir and ganciclovir (pyrimidine derivatives) and causes the accumulation of monophosphate derivatives of these drugs in the transduced malignant cells, which will ultimately inhibit DNA polymerase and causes cell death. Indeed, studies in animal models have shown a significant increase in survival time with intrathecal application of thymidine kinase containing herpes vectors (Kramm *et al*, 1996). Experimental models have shown that a number of non-transduced neighboring tumor cells are also destroyed. One explanation for this phenomenon, called the 'bystander effect,' may be the existence of gap junctions between tumor cells, allowing for the exchange of tumoricidal factors (Dilber *et al*, 1997). Other mechanisms including immune mediated destruction and inflammatory process may be involved. Similarly, cytosine deaminase, a bacterial and fungal enzyme, can deaminate 5-fluorocytosine to 5-fluorouracil (5-FU), which is a commonly used chemotherapeutic agent in cancer treatment (Ge *et al*, 1997). Introduction of a cytosine deaminase gene into the glioma tumor cells will cause the transduced cells to accumulate 5-FU, which inhibits thymidylate synthesis causing subsequent cell death.

Another experimental approach has been the introduction of genes into the tumor cells that code for cytokines, e.g., interleukin-2 (Breder *et al*, 1996), or antisense molecules against growth factors. The rationale for the use of IL-2 is that this molecule can mobilize a cytotoxic immune response against tumor cells given that the patient is sensitized to autologous tumor cells. In fact, applied in this manner, the treatment caused substantial tumor shrinkage as was shown by imaging studies (Sobol *et al*, 1995).

Gene transfer methods for treatment of malignant gliomas

The technology to effectively deliver the 'suicide' genes into the tumor cells, while sparing the normal brain tissue, is at its infancy. It suffers from the same

drawbacks as chemotherapy. Much like other therapies, for a cure to occur, all malignant cells must be effectively destroyed. A variety of viral vectors including adeno-associated (Mizuno *et al*, 1996), adenoviral (Boviatsis *et al*, 1994; Okada *et al*, 1996; Maron *et al*, 1996; Fueyo *et al*, 1996), retroviral (Takamyia *et al*, 1993; Short *et al*, 1990; Culver *et al*, 1992) as well as non-viral methods (Zhu *et al*, 1996) have been devised to carry the given genes into the cells. However, in the case of gliomas, the seemingly simple mechanics of effectively reaching a proportion of tumor cells to effect a cure has proven to be surprisingly difficult. Currently, little information is available regarding the efficacy of gene therapy for brain tumors both in terms of the number of tumor cells transduced by a given dose of genetic material, and the distribution of the transduced cells; i.e., the quantitative kinetics of gene therapy in general has not been worked out. Direct application of small and large molecules, e.g., nucleotides and active vectors into the tumor extracellular space by an infusion process, called convection enhanced delivery system (Kroll *et al*, 1996; Levy *et al*, 1997), is currently under evaluation (Neuwelt *et al*, 1994; Nilaver *et al*, 1995; Muldoon *et al*, 1995). This method circumvents the blood brain barrier, and may allow therapeutic molecules to reach not only the tumor core but also the advancing edge. Bradykinin-like drugs have been used to breach the blood brain barrier (Nilaver *et al*, 1995; Bartus *et al*, 1996; Elliot *et al*, 1996b) prior to selective intra arterial infusion of therapeutic agents including gene carrying vectors or modified viral particles. Modified viral particles can be used not only as vectors but as therapeutic agents. One such virus, a genetically engineered herpes simplex virus (gamma 34.5), capable of replicating in the dividing tumor cells but avirulent to the surrounding terminally differentiated normal tissue has been studied (Mineta *et al*, 1995; Boviatsis *et al*, 1994; Andreansky *et al*, 1996). Application of these viral particles to a xenogeneic mouse glioma model improved survival (Andreansky *et al*, 1996). Use of this type of viral therapy in human trials must be carefully considered because of their potential for producing lethal encephalitis.

Another approach is *ex vivo* gene therapy. This involves the *in vitro* engineering of cells capable of continuously producing anti-tumor factors. These factors would include, among others, suicide genes, cytokines (Lichter *et al*, 1995) and antisense molecules (Saleh *et al*, 1996). The engineered cells are implanted in the tumor bed and presumably the tumor killing effects would persist *in vivo*. The search for suitable donor cells and appropriate genes are under intense study. Allogeneic fibroblasts, lymphocytes, syngeneic normal and malignant glial cells, as well as xenogeneic cells, are being evaluated (Isacson and Breakfield, 1997).

Recently, we have been investigating the feasibility of using progenitor cells with astrocytic characteristics from adult brains, and marrow stromal cells (Prockop, 1997) as donor cells, and for cell mediated gene transfer into the nervous system (Azizi, 1997; Azizi *et al*, 1998). Astrocytes and bone marrow stromal cells, because of their post graft properties of migration (Anderson *et al*, 1993; Emmett *et al*, 1988; Lund *et al*, 1993), integration into the CNS (DelBigio *et al*, 1995) and elaboration of growth factors may be the ideal cells for *ex vivo* gene transfer.

Current molecular approaches

In recent years a wealth of information has emerged regarding the molecular mechanisms of regulation of the cell cycle and its role in the genesis of gliomas. One commonly known suppressor gene is p53, which is located in chromosome 17 and is involved in several aspects of cell cycle control as well as suppression of malignant transformation (Asai *et al*, 1994; Bogler *et al*, 1995). This may be accomplished either by inducing apoptosis or arrest in the cell cycle (Gomez-Manzano *et al*, 1996). Aberrant expression of the p53 gene is thought to be an early event in malignant transformation of many human astrocytic tumors (Haapasalo *et al*, 1993). It has been suggested that the loss of wild type p53 function is associated with genomic instability, accelerated growth and malignant transformation of cultured embryonic astrocytes (Yahanada *et al*, 1995). However, other studies have shown that the expression of wild type p53 may be increased in some malignant gliomas (Alderson *et al*, 1995; Bogler *et al*, 1995), thus indicating a complex molecular mechanism. One path for the involvement of p53 in the genesis of brain tumors may be enhancing the expression of p21, which in turn increases the phosphorylation activity of cyclin D1. A substrate for cyclin D1 is the retinoblastoma protein (pRB) (Mayol *et al*, 1995). This protein forms a suppressor complex with E2F. *E2F-1* is a ubiquitously expressed gene, and its product E2F-1 attaches to E2F promoter which augments the transcription of S-phase specific genes and orchestrates the transition from G1 to S-phase in the cell cycle (Parr *et al*, 1997). Thus, hyperphosphorylation

of pRB releases the E2F1 transcription factors, and accelerates the cell's progression to S-phase. It is possible that cells with 'defective' DNA, i.e., neoplastic cells, would accelerate through the S-phase and undergo apoptosis, whereas the malignant cells with defect(s) in this pathway would remain in G-phase for repair, become viable, proceed through the normal cell cycle and ultimately form brain tumors. Indeed, it has been reported that tumors that are resistant to radiotherapy have a faster repair mechanism of DNA breaks (Schwartz *et al*, 1988). It has been reported previously that a gene defect or loss of expression of pRB gene product in human malignant gliomas is associated with advanced disease (Hamel *et al*, 1993; Paggi *et al*, 1994; Dynlacht *et al*, 1997). Although the above pathway may be only one of the possibly many molecular mechanisms by which the genesis of brain tumors can occur, understanding of the steps in this pathway affords us a rich opportunity for therapeutic interventions at many points.

Another gene associated with gliomas is p16, located in 9p21 chromosomal region. Deletion of this region is one of the common chromosomal alterations during the evolution and genesis of gliomas. It has been observed that aberrations or deletions in p16 gene is associated with malignant transformation (Arap *et al*, 1997), whereas introduction/reintroduction of this gene into malignant glioma cells inhibit glioma proliferation (Fueyo *et al*, 1996; Hama *et al*, 1997) and induce the malignant cells into senescence (Uhrbom *et al*, 1997). Future studies on molecular therapies will be concentrated on harnessing the power of selectively targeted molecular therapeutics.

Conclusions

Despite a recent pessimistic report that the treatment of cancer over the past two decades has been a failure (Bailar and Gornik, 1997), major advances in understanding of the mechanisms of genesis of tumors has been made. These continued discoveries have inevitably brought us on the verge of launching new effective treatments for malignant gliomas.

(Supported by the Department of Neurology, In memory of Eileen S Kelberg).

References

- Alderson LM, Castleberg RL, Harsh GR IV, *et al* (1995). Human gliomas with wild-type p53 expresses bcl-2. *Cancer Research* **55**: 999–1001.
- Ameri A, Poisson M, Chauveinc L, *et al* (1997). Treatment of recurrent malignant supra tentorial gliomas with the association of carboplatin and etoposide: a phase II study. *J of Neuro-Oncology* **32**: 155–160.
- Anderson C, Tytell M, Brunso-Bechothold J (1993). Transplantation of cultured type-1 Astrocyte cell suspensions into young, adult and aged rat cortex: cell migration and survival. *Int J Dev Neuroscience* **11**: 555–568.

- Andreansky SS, He B, Gillespie GY, Soroceanu L, Markert J, Chou J, Roizman B, Whitley R (1996). The application of genetically engineered herpes simplex viruses to the treatment of experimental brain tumors. *Proc Natl Acad Sci USA* **93**: 11313–11318.
- Arap W, Knudsen ES, Wang JY, *et al* (1997). Point mutations can inactivate *in vitro* and *in vivo* activities of p16 (INK4A)/CDKN2A in human glioma. *Oncogene* **14**: 603–609.
- Asai A, Miyagi Y, Sugiyama A, *et al* (1994). Negative effects of wild-type p53 and s-Myc on cellular growth and tumorigenicity of glioma cells. Implication of the tumor suppressor genes for gene therapy. *J of Neuro-Oncology* **19**: 259–268.
- Azizi SA (1997). Adenovirus mediated gene transfer to astrocytes from adult rats: vehicles for *ex vivo* gene therapy. *J Neurovirology* **3** (suppl 1): S 73.
- Azizi SA, Stokes D, Augelli BJ, DiGirolamo C, Prockop DJ (1998). Engraftment and migration of human bone marrow stromal cells implanted in the brains of albino rats. Similarities to astrocyte grafts. *Proc Natl Acad Sci (USA)* (in press).
- Bailar JC, Gornik HL (1997). Cancer undefeated. *N England J Medicine* **336**: 1569–1574.
- Balmaceda C, Fetell MR, Hesdorffer C. (1997). Thiotepa and etoposide treatment of recurrent malignant gliomas: a phase I study. *Cancer Chemotherapy & Pharmacology* **40**(1): 72–74.
- Barth RF, AH Soloway (1997). Boron neutron capture therapy of brain tumors - current status and future prospects. *J Neuro-Oncology* **33**: 3–7.
- Barth RF, Yang W, Rotaru JH, Moeschberger ML, Joel DD, Nawrocky MM, Goodman JH, Soloway AH (1997). Boron neutron capture therapy of brain tumors: enhanced survival following intracarotid injection of either sodium borocaptate or boronophenylalanine with or without blood-brain barrier disruption. *Cancer Research* **57**: 1129–1136.
- Bartus RT, Elliott PJ, Dean RL, *et al* (1996). Controlled modulation of BBB permeability using the bradykinin agonist, RMP-7. *Experimental Neurology* **142**: 14–28.
- Beck-Bornholdt Hp, Dubben HH, Liertz-Peterson C, Willers H (1997). Hyperfractionation: Where do we stand? *Radiotherapy and Oncology* **43**: 1–21.
- Beutler AS, Banck MS, Aguzzi A, Wedekind D, Hedrich HJ (1997). Curing rat glioblastoma: Immune therapy or graft rejection? *Science* **276**: 20–21.
- Black PM (1997). Management of malignant Glioma: Role of surgery in relation to Multi modality Therapy. *J Neurovirology* (current issue).
- Blomgren H (1996). Brain Tumors. *Acta Oncologica* **35**(suppl 7): 16–21.
- Bodmer S, Strommer K, Fei K, Siepl C, deTribollet N, Heid I, Fontana A (1989). Immunosuppression and transforming growth factor beta in glioblastoma: preferential production of transforming growth factor beta-2. *J Immunol* **143**: 3222–3229.
- Bogler O, Huang HJ, Kleihues P, Cavenee WK (1995). The p53 gene and its role in human brain tumors. *GLIA* **15**(3): 308–327.
- Boiardi A, Silvana A, Pozzi A, *et al* (1997). Advantage of treating anaplastic gliomas with aggressive protocol combining chemotherapy and radiotherapy. *J of Neuro-Oncology* **34**: 179–185.
- Boring CC, Squires TS, Tong T, Montgomery S (1994). Cancer statistics. *Ca: a Cancer J for Clinicians* **44**: 7–26.
- Boviatsis EJ, Scharf JM, Chase M, Harrington K, Kowar NW, Breakfield Xo, Chiocca EA (1994). Antitumor activity and reporter gene transfer into rat brain neoplasms inoculated with herpes simplex virus vectors defective in thymidine kinase or ribonucleotide reductase. *Gene Therapy* **1**: 323–331.
- Brady L, Woo D, Markoe A, Dadvarpar S, Karsson U, Rackover M, Peyster R, Embrich J, Miyamoto C, Steplewski Z, Koprowski H (1990). Radioimmunotherapy with ¹²⁵I-EGF-425 in patients with brain tumors: Preliminary results of a phase II clinical trial. *Antibody Immunoconjugat Radiopharm* **3**: 169–179.
- Brandes AA, Rigon A, Zampieri P, *et al* (1996). Early chemotherapy and concurrent radio-chemotherapy in high grade glioma. *J of Neuro-Oncology* **30**: 247–255.
- Breder J, Ruller S, Ruller E, *et al* (1996). Induction of cell death by cytokines in cell populations restricted to G1 and G2. *Experimental Cell Research* **223**: 259–267.
- Chernauek SD (1993). Insulin-like growth factor I (IGF-I) production by astroglial cells: regulation and importance for epidermal growth factor-induced cell replication. *J of Neuroscience Research* **34**: 189–197.
- Culver KW, *et al* (1992). *In vivo* gene transfer with retroviral vector producer cells for treatment of experimental brain tumors. *Science* **256**: 1550–1552.
- Czubayko F, Liaudet-Coopman ED, Aigner A, *et al* (1997). A secreted FGF-binding protein can serve as the angiogenic switch in human cancer. *Nature Medicine* **3**: 1137–1140.
- Del Bigio MR, Colin C, Jacque CM (1995). Fine structure of astroglial integration into host brain following xenografting. *J Neuropath & Exp Neurol* **54**: 385–394.
- Dilber MS, Abedi MR, Christenson B, *et al* (1997). Gap junctions promote the bystander effect of herpes simplex virus thymidine kinase *in vivo*. *Cancer Research* **57**: 1523–1528.
- Dynlacht BD, Moberg K, Lees JA, *et al* (1997). Specific regulation of E2F family members by cyclin-dependent kinases. *Molecular & Cellular Biology* **17**: 3867–3875.
- Elliott TE, Dinapoli RP, O'Fallon JR, *et al* (1996a). Randomized trial of radiation therapy (RT) plus dibromodulcitol (DBD) versus RT plus BCNU in high grade astrocytoma. *J of Neuro-Oncology* **33**: 239–250.
- Elliott PJ, Hayward NJ, Dean RL, *et al* (1996b). Intravenous RMP-7 selectively increases uptake of carboplatin into rat brain tumors. *Cancer Research* **56**: 3998–4005.
- Emmett CJ, Lawrence JM, Seeley PJ (1988). Visualization of migration of transplanted astrocytes using polystyrene microspheres. *Brain Res* **447**: 223–233.
- Emrich JG, Bender H, Class R, Eshleman J, Miyamoto C, Brady LW (1996). *In vitro* evaluation of iodine-125-labeled monoclonal antibody (Mab 425) in human high-grade glioma cells. *American J of Clinical Oncology* **19**: 601–608.
- Ezzeddine CD, *et al* (1991). Selective killing of glioma cells in culture and *in vivo* by retrovirus transfer of herpes simplex virus thymidine kinase gene. *New Biologist* **3**: 608–614.

- Faillot T, Magdelenat H, Mady E, Stasiecki P, Fohanno D, Gropp P, Poisson M, Delattre J-Y (1996). A Phase I Study of an Anti-epidermal Growth Factor Receptor Monoclonal Antibody for the Treatment of Malignant Gliomas. *Neurosurgery* **39**: 478–483.
- Fine HA (1995). Novel biological therapies for malignant gliomas. Anti angiogenesis, immunotherapy, and gene therapy. *Neurologic Clinics* **13**: 827–846.
- Fountzilias G, Karavelis A, Makrantonakis P, et al (1997). Concurrent radiation and intra carotid cisplatin infusion in malignant gliomas: a feasibility study. *American J of Clinical Oncology* **20**: 138–142.
- Fowler JF (1984). What is next in fractionated radiotherapy? *British J Cancer (suppl.6)* **49**: S285–S300.
- Fried MP, Hsu L, Topulos GP, Jolesz FA (1996). Image-guided surgery in a new magnetic resonance suits: preclinical considerations. *Laryngoscope* **106**: 411–417.
- Fueyo J, Gomez-Manzano C, Yung WK, et al (1996). Adenovirus-mediated p16/CDKN2 gene transfer induces growth arrest and modifies the transformed phenotype of glioma cells. *Oncogene* **12**: 103–110.
- Fujiwara T, Matsumoto Y, Honma Y, et al (1995). A comparison of intra arterial carboplatin and ACNU for the treatment of gliomas. *Surgical Neurology* **44**: 145–150.
- Garrison JL, Berens ME, Shapiro JR, et al (1996). Interferon-beta inhibits proliferation and progression through S phase of the cell cycle in five glioma cell lines. *J of Neuro-Oncology* **30**: 213–223.
- Ge K, Xu L, Zheng Z, Xu D, Sun L, Liu X (1997). Transduction of cytosine deaminase gene makes rat glioma cells highly sensitive to 5-Fluorocytosine. *Int J of Cancer* **71**: 675–679.
- Gomez-Manzano C, Fueyo J, Kyritsis AP, et al (1996). Adenovirus-mediated transfer of the p53 gene produces rapid and generalized death of human glioma cells via apoptosis. *Cancer Research* **56**: 694–699.
- Good DJ, Polverini PJ, Rastinejad F, Le Beau MM, Lemons RS, Frazier WA, Bouck NP (1990). A tumor suppressor-dependent inhibitor of angiogenesis is immunologically and functionally indistinguishable from a fragment of thrombospondin. *Proc Natl Acad of Sci USA* **87**: 6624–6628.
- Gould S, Darzi A (1997). The magnetic resonance operating theater. *British J of Surgery* **84**: 595–597.
- Gupta N, Vij R, Haas-Kogan DA, et al (1996). Cytogenetic damage and the radiation-induced G1-phase checkpoint. *Radiation Research* **145**: 289–298.
- Haapasalo H, Isola J, Sallinen F, et al (1993). Aberrant p53 expression in astrocytic neoplasms of the brain: association with proliferation. *Amer J of Pathology* **142**: 1347–1351.
- Hall WA, Djalilian HR, Sperduto PW, et al (1995). Stereotactic radiosurgery for recurrent malignant gliomas. *J of Clinical Oncology* **13**: 1642–1648.
- Hama S, Sadatomo T, Yoshioka H, et al (1997). Transformation of human glioma cell lines with the p16 gene inhibits cell proliferation. *Anticancer Research* **17**: 1933–1938.
- Hamel W, Westphal M, Shephard HM (1993). Loss in expression of the retinoblastoma gene product in human gliomas is associated with advanced disease. *J Neuro-Oncology* **16**: 159–165.
- Hanahan D, Folkman J (1996). Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis. *Cell* **86**: 353–364.
- Harada K, Yoshida J, Mizuno M, et al (1995). Growth inhibition of intracerebral rat glioma by transfection-induced human interferon-beta. *J of Surgical Oncology* **59**: 105–109.
- Hayes RL, Koslow M, Hieseiger EM et al (1995). Improved long term survival after intracavitary interleukin-2 and lymphokine-activated killer cells for adults with recurrent malignant glioma. *Cancer* **76**: 840–852.
- Hochberg F, Pruitt A (1980). Assumptions in the radiotherapy of glioblastoma. *Neurology* **30**: 907–911.
- Hopkins K, Chandler C, Bullimore J, et al (1995). A pilot study of the treatment of patients with recurrent malignant gliomas with intratumoral yttrium-90 radio-immunoconjugates. *Radiotherapy Oncology* **34**: 121–131.
- Isacson O, Breakefield XO (1997). Benefits and risks of hosting animal cells in the human brain. *Nature Medicine* **3**: 964–969.
- Izquierdo M, Martin V, de Felipe P, Izquierdo JM, et al (1996). Human malignant brain tumor response to herpes simplex thymidine kinase (HSVtk)/ganciclovir gene therapy. *Gene Therapy* **3**: 491–495.
- Kelland LR, Edwards SM, Steel GG (1988). Induction and rejoining of DNA double-strand breaks in human cervix carcinoma cell lines of differing radiosensitivity. *Radiation Res* **116**: 526–538.
- Kim SH, Kim JH, Kolozsvary A, et al (1997). Preferential radiosensitization of 9L glioma cells transduced with HSV-tk gene by acyclovir. *J of Neuro-Oncology* **33**: 189–194.
- Kiu MC, Chang CN, Cheng WC, et al (1995). Combination chemotherapy with carmustine and cisplatin before, during and after radiotherapy for adult malignant gliomas. *J of Neuro-Oncology* **25**: 215–220.
- Kramm CM, Rainov NG, Sena-Esteves M, et al (1996). Long-term survival in a rodent model of disseminated brain tumors by combined intra-theal delivery of herpes vectors and ganciclovir treatment. *Human Gene Therapy* **7**: 1989–1994.
- Kroll RA, Pagel MA, Muldoon LL, Roman-Goldstein S, Neuwelt EA. (1996). Increasing volume of distribution to the brain with interstitial infusion: dose, rather than convection, might be the most important factor. *Neurosurgery* **38**: 746–752.
- Kruse CA, Roper MD, Kleinschmidt-DeMasters BK, et al (1997). Purified herpes simplex thymidine kinase Retrovector particles. *Cancer Gene Therapy* **4**: 118–128.
- Kuchelmeister K, Elborg B, Gullotta F (1995). Immunohistochemical detection of p53 protein in tumors of the central nervous system. *Pathologica* **87**: 498–502.
- Kyritsis AP, Yung WK, Jaeckle KA, et al (1996). Combination of 6-thioguanine, procarbazine, lomustine, and hydroxyurea for patients with recurrent malignant gliomas. *Neurosurgery* **39**: 921–926.
- Lai YS, Ramsay DA, Macdonald DR, Del Maestro RF (1997). Therapy-related chromosomal changes and cytogenetic heterogeneity in human gliomas. *J of Neuro-Oncology* **32**: 7–17.

- Lamond JP, Mehta MP, Boothman DA (1996). The potential of topoisomerase I inhibitors in the treatment of CNS malignancies: report of a synergistic effect between topotecan and radiation. *J Neuro-Oncology* **30**: 1–6.
- Levine AL, Sheline GE, Putin PH (1989). In *Cancer: principles and practice of oncology*. Davita et al (eds.) (Lippincott, Philadelphia) pp. 1557–1611.
- Levy RM, Ward S, Schalgeter K, Groothuis D (1997). Alternative delivery systems for antiviral nucleosides and antisense oligonucleotides to the brain. *J of Neuro-Virology* **3**(suppl.1): S74–S75.
- Libermann TA, Nusbaum HR, Razon N et al (1985). Amplification, enhanced expression and possible rearrangement of the EGF receptor gene in primary human brain tumors of glial origin. *Nature* **313**: 144–147.
- Liu L, Barth RF, Adams DM, Soloway AH (1995). Bispecific antibodies as targeting agents for boron neutron capture therapy of brain tumors. *J Hematology* **4**: 477–483.
- Lichter T, Glick RP, Kim TS, et al (1995). Prolonged survival of mice with glioma injected intra cerebrally with double cytokine-secreting cells. *J of Neurosurgery* **83**(6): 1038–1044.
- Liotta LA, Steeg PS, Stetler-Stevenson WG (1991). Cancer metastasis and angiogenesis: an imbalance of positive and negative regulation. *Cell* **64**: 327–336.
- Lorusso L, Nano R, Capelli E, et al (1994). Activated lymphocytes in glioblastoma: significance for anti-tumoral immunity. *Acta Neurologica* **16**: 198–205.
- Lund RD, Zhou H-F, Yee KT (1993). The migration of astrocytes after implantation to immature brains. *J Dev Neuroscience* **11**: 595–601.
- Maron A, Gustin T, Le Roux A, et al (1996). Gene therapy of rat C6 glioma using adenovirus-mediated transfer of the herpes simplex virus thymidine kinase gene: long-term follow-up by magnetic resonance imaging. *Gene Therapy* **3**: 315–322.
- Matsumoto Y, Fujiwara T, Nagao S (1995). Determinants of drug response in camptothecin-11-resistant glioma cell lines. *J Neuro-Oncology* **23**: 1–8.
- Mayol X, Garriga J, Grana X (1995). Cell cycle-dependent phosphorylation of the retinoblastoma-related protein p130. *Oncogene* **11**: 801–806.
- Mineta T, Rabkin SD, Yazaki T, Hunter WD, Martuza RL (1995). Attenuated multi-mutated herpes simplex virus-1 for the treatment of malignant gliomas. *Nature Medicine* **1**: 938–943.
- Miyamoto C, Brady LW, Rackover M, et al (1995). Utilization of ¹²⁵I monoclonal antibody in the management of primary glioblastoma multiforme. *Radiation Oncology Investigations* **3**: 126–132.
- Mizuno M, Yoshida J (1996). Tumor necrosis factor-alpha gene transfer agents anti-Fas antibody mediated apoptosis in human glioma cells. *Jap Journal of Cancer Research* **87**: 543–547.
- Muldoon LL, Nilaver G, Kroll RA, Pagel MA, Breakefield XO, Chiocca EA, Davidson BL, Weissleder R, Neuwelt EA (1995). Comparison of intracerebral inoculation and osmotic blood-brain barrier disruption for delivery of adenovirus, herpesvirus, and iron oxide particles to normal rat brains. *American J of Pathology* **147**: 1840–1851.
- Nakagawa H, Fujita T, Kubo S, et al (1994). Selective intra-arterial chemotherapy with a combination of etoposide and cisplatin for malignant gliomas: a preliminary report. *Surgical Neurology* **41**: 19–27.
- Nakagawa Y, Hatanaka H (1997). Boron neutron capture therapy. Clinical brain tumor studies. *J Neuro-Oncology* **33**: 105–115.
- Nakatsu S, Knodo S, Knodo Y, Yin D, Peterson JW, Kaakaji R, Morimura T, Kikuchi J, Barnett GH (1997). Induction of apoptosis in multi-drug resistant human glioblastoma cells by SN-38, a metabolite of the camptothecin derivative CPT-11. *Cancer chemotherapy and Pharmacology* **39**: 417–423.
- Natali PG, Nicotra MR, Bigotti, et al (1991). Comparative analysis of the expression of the extracellular-matrix protein tenascin in normal and human fetal, adult and tumor tissues. *Int J of Cancer* **47**: 811–816.
- Nehashi K, Yoshida J, Wakabayashi T, et al (1995). Growth inhibition of human glioma cells by super-induced human interferon-beta. *Neurologia Medico-Chirurgica* **35**: 719–722.
- Nilaver G, Muldoon LL, Kroll RA, Pagel MA, Breakefield XO, Davidson BL, Neuwelt EA (1995). Delivery of herpesvirus and adenovirus to nude rat intracerebral tumors after osmotic blood-brain barrier disruption. *Proceedings of Nat Academy of Sciences of USA* **92**: 9829–9833.
- Nelson DF, Curran WJ, Scott C, Nelson JS, Weinstein AS, Ahmad K, Constine LS, Murray K, Powlis WD, Mohiuddin M, Fischbach J (1993). Hyperfractionated radiation therapy and bis-chloroethyl nitrosurea in the treatment of malignant glioma. *Int J Radiation Oncology Biol Phys* **25**: 193–207.
- Neuwelt EA, Weissleder R, Nilaver G (1994). Delivery of virus-sized iron oxide particles to rodent CNS neurons. *Neurosurgery* **34**: 777–784.
- Ohta S, Yanagihara K, Nagata K (1997). Mechanism of apoptotic cell death of human gastric carcinoma cells mediated by transforming growth factor beta. *Biochemical Journal* **324**: 777–782.
- Okada H, Miyamura K, Itoh T, et al (1996). Gene therapy against an experimental glioma using adeno-associated virus vectors. *Gene Therapy* **3**: 957–964.
- Okamoto S, Yoshikawa K, Obata Y, Shibuya M, Aoki S, Yoshida J, Takahashi T (1996). Monoclonal antibody against the fusion junction of a deletion-mutant epidermal growth factor receptor. *British J of Cancer* **73**: 1366–1372.
- Oldfield EH, Ram Z, Culver KW, et al (1993). Gene therapy for the treatment of brain tumors using intratumoral transduction with the thymidine kinase gene and intravenous ganciclovir. *Human Gene Therapy* **4**: 39–69.
- Packer RJ (1996). Alternative therapies for children with brain stem gliomas: immunotherapy and gene therapy. *Pediatric Neurosurg* **24**: 217–222.
- Paggi MG, Martelli F, Fanciulli M, et al (1994). Defective human retinoblastoma protein identified by lack of interaction with the E1A oncoprotein. *Cancer Research* **54**: 1098–1104.
- Parr MJ, Manome Y, Tanaka T, Wen P, Kufe DW, Kaelin WG and HA Fine (1997). Tumor selective transgene expression *in vivo* mediated by an E2F responsive adenoviral vector. *Nature Medicine* **3**: 1145–1149.

- Patel K, Rossell RJ, Bourne S, Walsh FS, Kemshead JT (1989). *Int J Cancer* **44**: 1062–1068.
- Peters LJ, Withers HR, Thames HD (1982). Radiobiological bases for multiple daily fractionation. In *Progress in radio-oncology II*. Kaercher DH, Kogelnik HD, Reinartz G. (eds.) Raven Press: New York pp 317–323.
- Prados MD, Warnick RE, Mack EE, *et al* (1996). Intravenous carboplatin for recurrent gliomas. A dose-escalating phase II trial. *American J of Clinical Oncology* **19**: 609–612.
- Press OW, Vitetta ES, Farr AG, *et al* (1986). Evaluation of ricin A-chain immunotoxins directed against human T cells. *Cellular Immunology* **102**: 10–20.
- Prockop DJ (1997). Marrow stromal cells as stem cells for nonhematopoietic tissues. *Science* **276**: 71–74.
- Rampling R, Steward W, Paul J, Macham MA, Harvey E, Eckley D (1994). rhGM-CSF ameliorates neutropenia in patients with malignant glioma treated with BCNU. *British J of Cancer* **69**: 541–545.
- Reifenberger G, Prior R, Deckert M, Wechsler W (1989). epidermal growth factor receptor expression and growth fraction in human tumors of the nervous system. *Virchows Arch (A)* **414**: 147–155.
- Rubin P, Gash DM, Hansen JT, Nelson DF, Williams JP (1994). Disruption of blood-brain barrier as the primary effect of CNS irradiation. *Radiotherapy and Oncology* **31**: 51–60.
- Saleh M, Stacker SA, Wilks AF. (1996). Inhibition of growth of C6 glioma cells *in vivo* by expression of antisense vascular endothelial growth factor sequence. *Cancer Research* **56**: 393–401.
- Sandberg-Nordqvist AC, Stahlbom PA, Reinecke M, *et al* (1993). Characterization of insulin-like growth factor I in human primary brain tumors. *Cancer Research* **53**: 2475–2478.
- Schupack K, Malkin M, Anderson L, *et al* (1995). The relationship between the technical accuracy of stereotactic interstitial implantation for high grade gliomas and the pattern of tumor recurrence. *Int J of Radiation Oncology Biol Phys* **32**: 1167–1176.
- Schwartz JL, Rotmensch J, Giovanazzi S, *et al* (1988). Faster repair of DNA double-strand breaks in radio resistant human tumor cells. *Int J Radiation Oncology Biol Phys* **15**: 907–912.
- Shepherd SF, Laing RW, Gosgrove VP, Warrington AP, Hines F, Ashley SE, Brada M (1997). Hypofractionated stereotactic radiotherapy in the management of recurrent glioma. *Int J Rad Oncology Biol Phys* **37**: 393–398.
- Shoenberg BS (1983). In *Oncology of the nervous system*. Walker MD (ed.) (Nijhoff, Boston) pp. 1–30.
- Short MP, *et al* (1990). Gene delivery to glioma cells in rat brain by grafting of a retrovirus packaging cell line. *J of Neuroscience Res* **27**: 427–439.
- Shrieve DC, Loeffler JS (1995). Advances in radiation therapy for brain tumors. *Neurol Clinics* **13**: 773–793.
- Sipos EP, Tyler B, Piantadosi S, *et al* (1997). Optimizing interstitial delivery of BCNU from controlled release polymers for the treatment of brain tumors. *Cancer Chemotherapy & Pharmacology* **39**: 383–389.
- Sneed PK, McDermot MW, Gutin PH (1997). Interstitial brachytherapy procedure for brain tumors. *Seminars in Surg Oncology* **13**: 157–166.
- Sobol RE, Fakhrai H, Shawler D, *et al* (1995). Interleukin-2 gene therapy in a patient with glioblastoma. *Gene Therapy* **2**: 164–167.
- Tachibana I, Moto M, Adjei PN, Gores GJ, Subramaniam M, Spelsberg TC, Urrutia R (1997). Over expression of the TGF-beta regulated Zinc finger encoding gene, TIEG, induces apoptosis in pancreatic epithelial cells. *J of Clin Investigations* **99**: 2365–2374.
- Takamiya Y, *et al* (1993). An experimental model of retrovirus gene therapy for malignant tumors. *J of Neurosurgery* **79**: 104–110.
- Thames HD, Peters LJ, Withers HR, Fletcher GH (1983). Accelerated fractionation vs. hyperfractionation: rationales for several treatments per day. *Int J Radiation Oncology Biol Phys*. **9**: 127–138.
- Trojan J, Johnson TR, Rudin SD, *et al* (1993). Treatment and prevention of rat glioblastoma by immunogenic C6 cells expressing antisense insulin-like growth factor I RNA. *Science* **259**: 94–97.
- Tsugu A, Kijima H, Yamazaki H, Ohnishi Y, Takamiya Y, Abe Y, Ueyama Y, Sato O, Tamaoki N, Nakamura M (1997). Localization of aberrant messenger RNA of epidermal growth factor receptor (EGFR) in malignant glioma. *Anticancer Research* **17**: 2225–2232.
- Tuzi NL, Venter KJ, Kumar S, Staddon SL, Lemoine LR, Gullick WJ (1991). Expression of growth factor receptors in human brain tumors. *Br J Cancer* **63**: 227–233
- Uhm JH, Dooley NP, Villemure JG, Yong VO (1997). Mechanisms of glioma invasion: role of matrix-metalloproteinases. *Can J Neurological Sci* **24**: 3–15.
- Uhrbom L, Nister M, Westermarck B (1997). Induction of senescence in human malignant glioma cells by p16INK4A. *Oncogene* **15**: 505–514.
- Valtonen S, Timonen U, Toivanen P, *et al* (1997). Interstitial chemotherapy with carmustine-loaded polymers for high-grade gliomas: a randomized double-blind study. *Neurosurgery* **41**: 44–48.
- Van Tassel P, Bruner JM, Maor MH, *et al* (1995). MRI of toxic effects of accelerated fractionation radiation therapy and carboplatin chemotherapy for malignant gliomas. *American J of Neuroradiology* **16**: 715–726.
- Waldman T, Zhang Y, Dillehay L, *et al* (1997). Cell-cycle arrest versus cell death in cancer therapy. *Nature Medicine* **3**: 1034–1036.
- Wara WM, Edwards MSB, Levin VA *et al* (1986). A new treatment regimen for brainstem glioma: a pilot study of the Brain Tumor Research Center and Children's Cancer Study Group. *Int J of Radiation Oncology Biol Phys* **12**(suppl.1): 143–144.
- Warnick RE, Prados MD, Mack EE, *et al* (1994). A phase II study of intravenous carboplatin for the treatment of recurrent gliomas. *J of Neuro-Oncology* **19**: 69–74.
- Weidner N (1996). Angiogenesis in breast cancer. *Cancer Treatment Res*. **83**: 265–301.
- Wiener HL (1995). The role of growth factor receptors in central nervous system development and neoplasia. *Neurosurgery* **37**: 179–193.
- Weingart JD, Thompson RC, Tyler B, Colvin OM, Brem H (1995). Local delivery of the topoisomerase I inhibitor camptothecin sodium prolongs survival in the rat intracranial 9L gliosarcoma model. *Int J of Cancer* **62**: 605–609.

- Werner-Wasik M, Scott CB, Nelson DF, Gaspar LE, Murray KJ, Fischback JA, Nelson JS, Weinstein AS, Curran WJ (1996). Final report of a phase I/II trial of hyperfractionated and accelerated hyperfractionated radiation therapy with carmustin for adults with supratentorial malignant gliomas. Radiation Therapy Oncology Group Study 83-02. *Cancer* **77**: 1535–1543.
- Wikstrand CJ, McLendon RE, Friedman AH, Bigner DD (1997). Cell surface localization and density of the tumor-associated variant of the epidermal growth factor receptor, EGFRvIII. *Cancer Research* **57**: 4130–4140.
- Wlodek D, Hittelman WN (1987). The repair of double-strand DNA breaks correlates with radiosensitivity of L5178Y-S and L5178Y-R cells. *Radiation Res* **112**: 146–155.
- Yahanada AM, Bruner JM, Donehower LA, Morrison RS. (1995). Astrocytes derived from p53-deficient mice provide a multi-step *in vitro* model for development of malignant gliomas. *Molecular & Cellular Biology* **15**: 4249–4259.
- Yokoyama S, Ohishi N, Shamoto M, *et al* (1997). Isolation and expression of rat interferon beta gene and growth-inhibitory effect of its expression on rat glioma cells. *Biochemical & Biophysical Research Communications* **232**: 698–701.
- Yang W, Barth RF, Rotaru JH, Moeschberger ML, Joel DD, Nawrocky MM, Goodman JH, Soloway AH (1997). Boron-Neutron capture therapy of brain tumors: enhanced survival following intracrotid injection of sodium borocaptate with or without blood brain barrier disruption. *Int J Radion Oncology Biolog Physics* **37**: 663–672.
- Yu JS, Burwick JA, Dranoff G, Breakefield XO (1997). Gene therapy for metastatic brain tumors by vaccination with granulocyte-macrophage colony-stimulating factor-transduced tumor cells. *Human Gene Therapy* **8**: 1065–1072.
- Zhu J, Zhang L, Hanisch UK, Felgner PL, Reszka R (1996). Continuous intracerebral gene delivery system for *in vivo* liposome-mediated gene therapy. *Gene Therapy* **3**: 476–476.